



Methodology for Health Monitoring of Mice Maintained in IVCs

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1



Health Monitoring (HM)

Pathogens/Subclinical Infections
 Opportunists, Pathobionts

Standardization:
 Definition of the Hygienic Status,
 Research Validity

Individual Study Confounders
 Microbiome

Barrier 1
 Barrier 2

Animal Health:
 Exclusion of Pathogens/Clinical Disease

Biosafety:
 Exclusion of Zoonotic Agents

Segmented Filamentous Bacteria
 Proteus sp.
 Helicobacter hepaticus
 Pneumocystis murina
 Ectromelia virus/mousepox
 Mouse Rotavirus
 Streptococcus moniliformis

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Bald et al. 2011, Laboratory Animals 2011; 45: 278-279. DOI: 10.1258/la.2010.010150
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Open Forum Infect Dis, 2017 Mar 14(2) e14028. https://doi.org/10.1093/ofid/ofw028
Editor, iCollection 2017 Spring
Milner et al. 2002, Infection Control and Hospital Epidemiology 2002; 27: 1022-1029. doi: 10.1017/S0950268802002025
Shen et al. 2020, Front Microbiol 2020; 11: 562348. doi: 10.3389/fmicb.2020.02025

4



Tasks of the Working Group (WG)

- Give recommendations for the health monitoring (HM) of mice maintained in IVCs

Recommend practical definitions of microbiological units for IVC husbandry

Review sampling and detection methods

Evaluate different strategies with special regard to the 3R's and research validity

Discuss advantages and disadvantages of different samples and detection methods

Support persons responsible for HM programs to establish and adapt their existing HM

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2



Definition of the agents

Panel of agents tested have to be individually defined, based on:

- Existing FELASA recommendations (which is not an exclusion list!)
- Risk of agent introduction and relevance of agents: Individual considerations
 - Facility management/purpose of units/immune status of animals
 - Improvement of the microbiological quality during the last decades
 - Relevance of commensal bacteria and the microbiome
- Recent Updates
 - Changes in nomenclature/reclassifications (e.g. *Pasteurellaceae*)
 - Novel agents (e.g. Mouse Kidney Parvovirus, Murine Astrovirus)

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Milner et al., 2024, Laboratory Animals, 2024;48(3): 278-292

5



Members of the WG

- Convenor
 Isabelle Goncalves da Cruz (AFSTAL)
- Members
 Marion Berard (AFSTAL)
 Ferdinando Scavizzi (AISAL)
 Stephanie Buchheister (GV-SOLAS)
 Arthur Humbert (SGV)
- Corresponding member
 Lorna Cleverley

AFSTAL
 Association Française pour l'Étude et le Soins des Animaux de Laboratoire

AISAL
 Associazione Italiana per lo Studio e la Cura degli Animali da Laboratorio

GV-SOLAS
 Gesellschaft für Versuchstierkunde
 Society for Laboratory Animal Science

SGV
 Swiss Laboratory Animal Science Association

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3



Individually Ventilated Cages (IVCs)

- Equipment is designed to prevent (reduce) the spreading of infectious agents
- Challenges:**
 - Definition of microbiological units: **Sentinel animal** based test strategies
- Require efficient transmission of agents: Reduced diagnostic sensitivity

Used Cage/ Dirty Bedding Sentinels
 Colony
 Contact sentinels
 Exhaust air sentinels
 FILTER

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 Doctoral Thesis Lena Brix, TiHo Hannover, 2023
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6

Environmental Sampling Strategies

➤ Molecular methodology enables the use of environmental sampling strategies

e.g. cage feces, cage swabs, bedding/nesting material, exhaust air dust (EAD)



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7

Optimizing sampling



- **False positive and false negative results**
- **Quality of samples, number of samples**
When **pooling**, keep in mind:
Pathogen dilution might cause false negative results
Only reasonable for animals that are representative from the same microbiological unit
Seek advice from the testing lab to match the requirement of diagnostic methods
= **balance between cost and quality**
- **Reduction or Replacement** of animals; **Refinement** and welfare of sentinels
- **Storage and transport of samples**

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<https://www.felasa.com/publications/doi/10.24452/23081>
<https://publicdomainpictures.org/file/stock-photo-velo-borgstrassen-LKW-Vektor-sfz0urfa/2019.html>

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10

Comparison of Strategies

100% animals sentinels + colony animals 	Mix of animals and environmental 	100% environmental
use of a large number of dedicated animals		no animals used
questions of suitability of strains (outbred vs. inbred vs. immunodeficient)		no questions about strains
various diagnostic methodology	Good compromise? Complex strategies required!	molecular methodology only
diagnostic sensitivity varies		improved diagnostic sensitivity
all known agents can be tested		not validated for all agents, yet
control for false negatives		control for false positives

Always perform follow up diagnostic of sick animals

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8

About the results

- What to do when there is a positive?
 - Confirm result – re-testing
infectious agents vs. residual nucleic acids
 - Isolate the contaminated microbiological unit quarantine measures
 - Decide about the fate of the colony
re-derivation vs. termination
- **Communication of the results!**
Proper description of HM concepts and methodology in health reports
Suspicious results should be reported, comments on measures taken



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<https://www.publicdomainpictures.net/de/view-image.php?image=65515&picture=glass-of-the-red-wine>

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11

Choice of the best method (for each agent)

Agent	Animal/Sentinels 	Method	Environmental 	Method
Murine Norovirus	Yes, bedding sentinels, Dubelko 2018 Yes, bedding sentinels, Hanson 2021 No, bedding sentinels, Miller 2018 Yes, bedding sentinels, O'Connell 2021 Less efficient, bedding sentinels, Zorn 2016	Serology PCR	Yes, filters, Dubelko 2018 Yes, media in soiled bedding, Hanson 2021 Yes, filters, O'Connell 2021 No, exhaust debris, Bauer 2016 Yes, EAD, Pettan-Brewer 2020 Yes, EAD, Zorn 2016	PCR (NGS)
Rodentibacter sp.	Yes, bedding sentinels, Dubelko 2018 No, bedding sentinels, Miller 2018 Less efficient, bedding sentinels, Miller 2016 Yes, bedding sentinels, Roepener 2018	Culture PCR	Yes, filters, Dubelko 2018 Yes, media in soiled bedding, Bauer 2016 Yes, EAD, Mahabir 2019 Yes, EAD, Miller 2016	PCR (NGS)
Ectoparasites	Yes, bedding sentinels, Gerwin 2017 No, bedding sentinels, Hanson 2021 Yes, bedding sentinels, De Bruin 2016 No, bedding sentinels, Miller 2018 Yes/No, bedding sentinels, Körner 2019	Microscopy PCR	Yes, Filter top, Gerwin 2017 Yes, media in soiled bedding, Hanson 2021 Yes, EAD, Körner 2019 Yes, exhaust debris, Bauer 2016	PCR (NGS)

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9

Summary

- HM of mice maintained in IVC is **challenging**
- **Microbiological units** have to be defined based on husbandry practices and sampling approaches
- **Environmental sampling strategies** are used to improve animal welfare and diagnostic sensitivity
- Environmental sampling strategies **have limitations** and require **critical result interpretation**
- Optimize sampling procedures and do "not forget the animals!"
- Timeline: Currently writing of the paper, **publication expected early 2024**

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12

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Thank you!

 **SGV** swiss laboratory animal science association

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 **GV-SOLAS**
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